Hybridization Buffer (20mL, in a 50 mL Falcon tube)
2 g Dextran sulfate (lives in the cabinet above the incubation oven)
3.6 mL NaCl 5M (lives in 50 mL Falcon tube on bench)
400 µL Tris-HCl 1M (lives in 50 mL Falcon tube on bench)
Combine, warm and dissolve in water bath at 55 ℃ for 2-3 hours vortexing every hour
Once dissolved cool down to 4 ℃ (room temp)
__In the 2-3 hours take out Blocking Reagent aliquot from freezer to thaw__
8 mL of Formamide (in small amber bottle in shelf in the doorway of the lab fridge)
2 mL 10% Blocking Reagent (found in skinny Falcon tubes in door of lab freezer)
6 mL MiliQ water
200 µL SDS 1% (10 µL added, stock is @ 20 %, stock lives in 50 mL Falcon tube on bench)

Store @ -20 ℃ for up to a year, label tube “ALV01 Hyb. Buffer, 40% Formalin, Date made)

Wash Buffer (made each time hybridization is conducted in graduated cylinder)
50 mL for 6 samples, 100 mL for 12 samples
1 mL Tris-HCl 1M
460 µL NaCl 5M
500 µL EDTA 0.5 M
25 µL SDS 20%
Bring up to 50 mL with DNA or RNA clean MiliQ water in hood
*if doing 12 samples double all reagent amounts including MiliQ water

TNT Buffer (1L)
In a large graduated cylinder add:
100 mL Tris-HCl 1M
30 mL NaCl 5M
Bring up to 1 mL with MilliQ water
Add 740 µL Tween (found on bench top, very viscous)
Invert jar to mix
Store @ room temp and label with new date it was made

PI/Citifluor Mounting Reagent
In a 1.5 mL eppendorf tube
1 mL Citifluor AF1 (in bin in lab fridge, clear and very viscous)
10 µL Propidium Iodide (in amber plastic bottle in bin in lab fridge, stains nucleus red)